Structure-function analysis of neuroprotectants

In "<u>The chemistry of neuroprotection</u>", the author argues convincingly that there could be great benefit from a systematic and rigorously scientific study of the physical chemistry of putative neuroprotectants vis-à-vis their pharmacological effect. However, the first example used of the earliest thinking in this direction (which comes, not surprisingly via V. A. Negovskii, the father of resuscitation¹ medicine) is instructive as to some of the potential barriers standing in the way of this approach.

"It is not surprising that all the agents which are effective in shock carry a negative charge. This applies both to heparin, which possesses a very strong negative charge, and to hypertonic glucose solution. The same may be said about a substance now in wide use - dextran - which has small, negatively charged molecules, and also about the glucocorticoids 21, 17, and 11, which also have a negative charge." - Professor Laborit in: Acute problems in resuscitation and hypothermia; proceedings of a symposium on the application of deep hypothermia in terminal states, September 15-19, 1964. Edited by V. A. Negovskii.

In the intervening decades since Laborit wrote the words guoted above, supraphysiologic (high) steroids have not only failed to demonstrate benefit in cerebral resuscitation and shock, they have been found to be actively harmful in every well designed RCT undertaken to test their utility (a). This also extends to their lack of utility in trauma, spinal cord injury and sepsis. Similarly, the utility of heparin in treating the encephalopathy of the post-resuscitation syndrome, or improving survival after cardiac arrest has recently been called into question. Glucose, hypertonic or otherwise, was long ago demonstrated to markedly increase neurological injury if given immediately after reperfusion following cardiac arrest, and elevated blood levels of glucose, both pre- and post cardiac arrest have a strong negative correlation with both survival neurological outcome. and

Determining the seriously harmful effects of steroid administration in critical illness took decades. Despite the compelling evidence for their injurious effects, administration of large, supraphysiologic doses of steroids is still a practice both used and defended by some clinicians (albeit not ones who rely on evidence based criteria) and the use of glucose in shock, trauma and cardiac arrest took a nearly comparable period of time to discredit. These two examples are noteworthy because they comprised mainstays of therapy for most kinds of neuroinjury for decades, and they had compelling theoretical appeal, as well as many positive small clinical and animal research studies. Indeed, the debate continues to this day with controversy centred mostly on the use of low or "physiological replacement" doses of steroids in

¹ Resuscitation medicine is properly termed *reanimatology*, and is so-called in the non-English speaking world

critical illness. As the eminent pulmonologist and intensivist Neil Macintyre observed in 2005, "Patients die, but steroids never do." This raises the twin problems of bad research (i.e., junk science) and statistically under powered or otherwise flawed studies. Combined, it has been estimated that these two types of defective studies comprise the bulk of published peer-reviewed scientific work.

High dose corticosteroid therapy for neuroinjury offers another complication in determining the therapeutic efficacy of any drug that merits consideration as a neuroprotectant (new or old). While there is no doubt that high-dose corticosteroids are ineffective and deleterious in the clinical setting, there is also little doubt that these agents *are* neuroprotective in the laboratory setting under certain conditions and for discrete subpopulations of neurons. The reasons for the failure of translational research in the case of corticosteroids are complex, but are mostly attributable to crucial differences between the laboratory and the real world of clinical medicine. In the case of corticosteroids these differences are most significantly:

- a. Delay from time of insult to time of treatment; in the laboratory the timing of interventions is uniform and is typically much shorter than is the case in the clinic where delays in both presentation and treatment are both long and highly variable.
- b. Heterogeneity of injury in humans compared to animals; animal models of neuroinjury are highly standardized (location, extent, mechanics) whereas human patients present with diverse injuries inflicted in many complex and often poorly understood ways.
- c. Species differences; not only are there large genetic differences between humans and rodents in general, there are dramatic differences in the native ability of rodents to both resist and overcome infection in comparison to humans.
- d. Demographics and comorbidities: laboratory animals are comparatively very uniform genetically, are typically young and healthy and of the same age, do not have comorbid conditions such as hypertension, diabetes, atherosclerosis, obesity or the diminished physiological capacity and repair and regenerative capacity increasingly present in humans over the age of 25.
- e. Rodents aren't people and do not interact with investigators in ways that facilitate straightforward determination of an adverse affect such loss of short term memory, or other cognitive deficits. It is now understood that the corticosteroids are toxic to the neurons of the hippocampus in both rodents and men. However, injury from this adverse effect is not only more evident in men than in mice (or rats for that matter), it is only men who are capable of complaining about it.

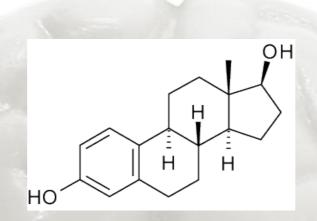
It is notable that all of these effects, with the possible exception of increased resistance to steroid-induced immunosuppression-mediated infection, obtain in the case of other translational models of drug development. The conclusion that corticosteroids are very likely neuroprotective in humans (in terms of the direct pharmacological effect on selected subpopulations of neurons in injured central nervous tissues under ideal conditions) is highly likely. However, the confounding

realities of the clinic and the genetic differences between men and rodents (the animals almost exclusively used in this type of research) mask this effect. This poses yet another serious challenge to investigators seeking to establish common moieties in prospective neuroprotective molecules.

Clinical trials of putative neuroprotective substances have been overwhelmingly negative. This has been the outcome despite often stellar results achieved in animal models; often in diverse species in studies conducted by multiple investigators in different institutions and sometimes in different countries; none of whom have any obvious relationship, let alone one that might raise the specter of conflict of interest. In the last 6 years alone, over 1000 experimental papers and over 400 clinical articles have appeared on this subject. What this suggests is that the same deficiencies seen in studies reported upon in rest of the peer-reviewed biomedical literature also apply to studies of pharmacological intervention in neuroprotection. An inevitable conclusion is that until the signal to noise ratio improves, attempts to draw general conclusions about the shared, essential properties of neuroprotective molecules will be difficult at best, and unreliable or misleading at worst.

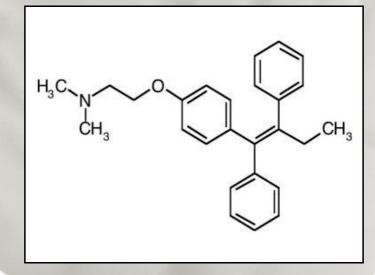
Perhaps a good place to start this kind of analysis is in an area where the molecular structure of the agent(s) is extraordinarily simple and the animal and clinical data are both robust and show good to fair agreement. Hypertonic sodium chloride solutions have demonstrated efficacy in providing both systemic (splanchnic) and cerebral protection in a broad class insults including hemorrhagic/hypovolemic shock, closed head injury and less robustly in stroke and global cerebral ischemia. Interestingly, other cation salts of chloride given at comparably high tonicity do not have this effect. Furthermore, animal as well as small human clinical studies have demonstrated isochloremic hypertonic solutions to be as effective as hypertonic sodium chloride at restoring microcirculatory flow and reversing metabolic acidosis in haemorrhagic shock without the potentially troublesome side-effect of raising the mean arterial pressure to levels where re-bleeding may occur in trauma or subarachnoid haemorrhage. A relative lack of effectiveness of the chloride salt of magnesium compared to the sulfate salt of this ion has also been noted. Understanding the mechanics of these paradoxes would seem to be a worthwhile and comparatively straightforward place to begin such structure-activity relationship analyses.

17β-Estradiol



Cerebroprotective drugs not infrequently possess a multiplicity of pharmacological effects that are known to be neuroprotective but that may be accomplished by very different and even indirect means in terms of their structure-function relationship. Some cerebroprotective molecules, such as the female hormone 17β-estradiol and the mixed estrogen antagonist-agonist tamoxifen share common physiochemical properties such as free radical scavenging, N-methyl-d-aspartate (NMDA) receptor inhibition, and modulation of volume regulated anion channels (VRAC); which play a role in ischemia-induced release of excitatory amino acids. There is considerable evidence that some of 17β-estradiol's neuroprotective effect is via signal transduction as well as its neurotrophic effects, even at doses below those necessary for its direct effects on reactive oxygen species production and its NMDA receptor inhibiting effects. While the structure of the molecules shares some important features, they are also structurally very different and the signal transduction and neurohormonal effects are almost certainly very different. Thus, these molecules also present a fascinating opportunity to probe structure-function relationships in neuropharmacology.

Tamoxifen



Finally, an admission, or perhaps a confession is order in ending this discussion. This author has been responsible for the application of at least one putative neuroprotective drug to cryopatients which ultimately proved ineffective in human clinical trials when administered during and after cardiopulmonary resuscitation (CPR). This drug, nimodipine, performed well in animal trials, but failed to show benefit in human trials, possibly as a result of its hypotension-inducing effect. Adequate mean arterial pressure (MAP) following resuscitation from cardiac arrest is essential to survival and a post arrest bout of hypertension has been demonstrated to provide substantial cerebral rescue in animal models of global cerebral ischemia. Reduction of MAP in cryopatients is a serious concern because achieving adequate perfusion pressure is problematic under the best of conditions. It is also worth noting that cryopatients have been given a variety of other ineffective neuroprotective drugs over the past 30 years, including the opiate agonist naloxone, the corticosteroid methylprednisolone and the iron chelating drug desferroxamine.

While these drugs, with the possible exception of nimodipine, are not likely to have been injurious (except perhaps to the pocketbook), their use raises important questions about when and how promising animal research should be translated to the setting of clinical cryonics. Unique among all other populations of human and animal patients, cryopatients have the opportunity to be treated with neuroprotective drugs that show great promise, absent the long delays of regulatory vetting, and independent of the economic pressure experienced by pharmaceutical companies to not only market drugs that are effective, but to market ones that are also profitable. The question thus becomes what criteria do we use in applying these drugs absent the extensive pre- and post marketing evaluation that obtains with approved ethical drugs? In essence the question we must ask and answer is "can we do better, much better in fact, than our colleagues in conventional critical care medicine?

(a) The one condition in which there is unequivocal benefit to supraphysiologic administration of steroids is meningococcal meningitis with substantial evidence also supporting a similar degree of efficacy in Typhoid and Pneumocystis carinii pneumonia..

Michael G. Darwin, Independent Critical Care Consultant, <u>m2darwin@googlemail.com</u>

Selected Bibliography on Clinical Outcome with Corticosteroid Therapy in Pneumococcal Meningitis, Typhoid and PCP Pneumonia: Hoffman SL, Punjabi NH, Kumala S, et al: Reduction in mortality in chloramphenicol treated severe typhoid fever by high dose dexamethasone. N Engl J Med, 1984; 310:82-88.

Lebel MH, Freij BJ, Syrogainnopolous GA, et al: Dexamethasone therapy for bacterial meningitis: Results of two double-blind placebo-controlled trials. N Engl J Med, 1988; 319:964-971.

Montaner JS, Lawson LM, Levitt N, et al: Corticosteroids prevent early deterioration in patients with moderately severe Pneumocystis carinii pneumonia in the acquired immunodeficiency syndrome. Ann Intern Med, 1990;113:14-20.

Selected Bibliography on Clinical Outcome with Corticosteroid Therapy in Sepsis and Shock :

Bone RC, Fisher CJ Jr, Clemmer TP, et al: Early methylprednisolone treatment for septic syndrome and the adult respiratory distress syndrome. Chest, 1987; 92:1032-1036.

Cooperative Study Group, Bennet IL, Finland M, et al: The effectiveness of hydrocortisone in the management of severe infections. JAMA, 1963; 183:462-465.

Luce JM, Montgomery AB, Marks JD, et al: Ineffectiveness of high dose methylprednisolone in preventing parenchymal lung injury and improving mortality in patients with septic shock. Am Rev Respir Dis, 1988; 138:62-68.

The Veterans Administration Systemic Sepsis Cooperative Study Group: Effect of high-dose glucocorticosteroid therapy on mortality in patients with clinical signs of systemic sepsis. N Engl Med, 1987; 317:659-665.

Lucas C, Ledgerwood A: The cardiopulmonary response to massive doses of steroids in patients with septic shock. Arch Surg, 1984; 119:537-541.

Cronin L, Cook, DJ, Carlett J, Heyland DK, King D, Lansang MAD, Fisher JD Corticosteroid treatment for sepsis: A critical appraisal and meta-analysis of the literature. Critical Care Medicine, 1995;23(8):1438-1441.

Keh D, Sprung CL. Use of corticosteroid therapy in patients with sepsis and septic shock: an evidence-based review. Crit Care Med, 2004;32(11 Suppl:S527-33.

Lefering R, Neugebauer, EA. Steroid controversy in sepsis and septic shock: A meta-analysis. **Critical** Care Medicine, 7;1995;1293-1303.

Minneci PC, Deans KJ, Banks SM, Eichacker PQ, Natanson C. Meta-analysis: the effect of steroids on survival and shock during sepsis dependson the dose. Ann Intern Med, 2004:6;141:47-56.

Schumer W: Steroids in the treatment of clinical septic shock. Ann Surg, 1976; 184:333-341

Slotman G, Fisher C, Bone R: Detrimental effects of high dose methylprednisolone sodium succinate on serum concentration of hepatic and renal function indicators in severe sepsis and septic shock. Crit Care Med, 1993; 21:191-195

Zimmerman JJ. A history of adjunctive glucocorticoid treatment for pediatric sepsis: moving beyond steroid pulp fiction toward evidence-based medicine. Pediatr Crit Care Med, 2007;8:530-9.

Selected Bibliography on Clinical Outcome with Corticosteroid Therapy in Spinal Cord Injury Ischemic Stroke and Head Trauma:

Coleman WP, Benzel D, Cahill DW, Ducker T, Geisler F, Green B, Gropper MR, Goffin J, Madsen PW 3rd, Maiman DJ, Ondra SL, Rosner M, Sasso RC, Trost GR, Zeidman S. A critical appraisal of the reporting of the National Acute Spinal Cord Injury Studies (II and III) of methylprednisolone in acute spinal cord injury. J Spinal Disord, 2000;13:185-99.

CRASH Trial Collaborators, Final Results of MRC CRASH, a randomised placebocontrolled trial of intravenous corticosteroid in adults with head injury – outcomes at 6 months. Lancet, 2005;365:1957-59

Methylprednisolone for acute spinal cord injury: an inappropriate standard of care. [J Neurosurg. 2000].

Neurological and functional status 1 year after acute spinal cord injury: estimates of functional recovery in National Acute Spinal Cord Injury Study II from results modeled in National Acute Spinal Cord Injury Study III. [JNeurosurg. 2002].

Pointillart v, Petitjean ME, Wiart L, Vital JM, Lassie P, Thicoipe M, Dabadie P. Pharmacologic Therapy of spinal cord injury during the acute phase. Spinal Cord, 2000;38:71-76.

Qizilbash N, Lewington SL, Lopez-Arrieta JM. Corticosteroids for acute ischaemic stroke. Cochrane Database Syst Rev. 2000;(2):CD000064. Review. Update in: Cochrane Database Syst Rev. 2002;(2):CD000064.

Detrimental Effects of Hyperglycemia Pre- and Post-Insult in Regional and Global Cerebral Ischemia:

Anderson RE, Tan WK, Martin HS, Meyer FB. Effects of glucose and PaO2 modulation on cortical intracellular acidosis, NADH redox state, and infarction in the ischemic penumbra. Stroke, 1999;30:160–170.

Bruno A, Biller J, Adams HP Jr, Clarke WR, Woolson RF, Williams LS, Hansen MD, for the Trial of ORG 10172 in Acute Stroke Treatment (TOAST) Investigators. Acute blood glucose level and outcome from ischemic stroke. Neurology, 1999;52:280–284.

Bruno A, Levine SR, Frankel MR, Brott TG, Lin Y, Tilley BC, Lyden PD, Broderick JP, Kwiatkowski TG, Fineberg SE, and the NINDS rt-PA Stroke Study Group. Admission glucose level and clinical outcomes in the NINDS rt-PA Stroke Trial. Neurology, 2002;59:669–674.

de Courten-Myers G, Myers RE, Schoolfield L, Hyperglycemia enlarges infarct size in cerebrovascular occlusion in cats. Stroke, 1988; 19: 623 - 630.

de Courten-Myers GM, Kleinholz M, Wagner KR, Myers RE. Normoglycemia (not hypoglycemia) optimizes outcome from middle cerebral artery occlusion. *J Cereb* Blood Flow Metab, 1994;14:227–236.

Dietrich WD, Alonso O, Busto R. Moderate hyperglycemia worsens acute Els T, Klisch J, Orszagh M, Hetzel A, Schulte-Monting J, Schumacher M, Lucking CH. Hyperglycemia in patients with focal cerebral ischemia after intravenous thrombolysis: influence on clinical outcome and infarct size. Cerebrovasc Dis, 2002;13:89–94.

Kamada H, Yu F, Nito C, Chan PH. Influence of Hyperglycemia on Oxidative Stress and Matrix Metalloproteinase-9 Activation After Focal Cerebral Ischemia/Reperfusion in Rats: Relation to Blood-Brain Barrier Dysfunction. Stroke, 2007; 38: 1044 - 1049.

Kawai N, Keep RF, Benz AL. Hyperglycemia and the vascular effects of cerebral ischemia. Stroke, 1997;28:149–154.

Koistinaho J, Pasonen S, Yrjanheikki J, Chan PH. Spreading depression---induced gene expression is regulated by plasma glucose. Stroke, 1999;30:114–119.

Kraft SA, Larson CPJ, Shuer LM, Steinberg GK, Benson GV, Pearl RG. Effect of hyperglycemia on neuronal changes in a rabbit model of focal cerebral ischemia. Stroke, 1990;21:447–450.

Li PA, Shuaib A, Miyashita H, He QP, Seisjo BK. Hyperglycemia enhances extracellular glutamate accumulation in rats subjected to forebrain ischemia. Stroke, 2000;31:183–192.

Osuga S, Hogan MJ. In vivo uptake of [3H] nimodipine in focal cerebral ischemia: modulation by hyperglycemia. J Cereb Blood Flow Metab, 1997;17:1057–1065.

Parsons MW, Barber PA, Desmond PM, Baird TA, Darby DG, Byrnes G, Tress BM, Davis SM. Acute hyperglycemia adversely affects stroke outcome: a magnetic resonance imaging and spectroscopy study. Ann Neurol, 2002;52:20–28.

Prado R, Ginsberg MD, Dietrich WD, Watson BD, Busto R. Hyperglycemia increases infarct size in collaterally perfused but not end-arterial vascular territories. J Cereb Blood Flow Metab, 1988;8:186–192.

Puisinalli WA, Waldman S, Rawlinson D, Plum F. Moderate hyperglycemia augments ischemic brain damage: a neuropathologic study in rats. Neurology, 1982;32:1239–1246.

Rezkalla RH, Kloner RA. No-reflow phenomenon. Circulation, 2002; 105:656–662.

Ribo M, Molina C, Montaner J, Rubiera M, Delgado-Mederos R, Arenillas JF, Quintana M, Alvarez-Sabín A, Acute Hyperglycemia State Is Associated With Lower tPA-Induced Recanalization Rates in Stroke Patients. Stroke, 2005; 36: 1705 - 1709.

Sieber FE, Koehler RC, Brown PR, Eleff SM, Traystman RJ, Diabetic chronic hyperglycemia and cerebral pH recovery following global ischemia in dogs. Stroke, 1994; 25: 1449 - 1455.

Tsuda, K. Role of Hyperglycemia and Glutamate Receptors in Ischemic Injury in Acute Cerebral Infarction. Stroke, 2006; 37: 2199.

Wass CT, Lanier WL. Glucose modulation of ischemic brain injury: review and clinical recommendations. Mayo Clin Proc, 1996;71:801-12. Review.

Weir CJ, Murray GD, Dyker AG, Lees KR. Is hyperglycaemia an independent predictor of poor outcome after acute stroke? Results of a long term follow up study. BMJ, 1997;314:1303–1306.

A Few of the Many Negative Clinical Trials of Novel Neuroprotectants:

American Nimodipine Study Group, Clinical trial of nimodipine in acute ischemic stroke. The American Nimodipine Study Group. Stroke, 1992; 23, 3e8.

Calabrese EJ. Drug therapies for stroke and traumatic brain injury often display Ushaped dose responses: occurrence, mechanisms, and clinical implications. Crit Rev Toxicol, 2008;38(6):557-77. Review.

Cheng JD, Al-Khoury L, Zivin JA. Neuroprotection for Ischemic Stroke: Two Decades of Success and Failure. NeuroRX, 2004;1:36-45.

De Keyser J, Sulter **G**, Luiten PG. Clinical trials with neuroprotective drugs in acute ischaemic stroke: are we doing the right thing? Trends in Neurosciences, 1999;22: 535-540.

DeGraba TJ, Pettigrew C. Why do neroprotectivedrugs work in animals but not in humans? Neurologic Clinics, 2000;18:475-493.

Diener HC, Lees KR, Lyden P, Grotta J, Davalos A, Davis SM, Shuaib A, Ashwood T, Wasiewski W, Alderfer V, Hårdemark HG, Rodichok L; SAINT I and II Investigators.NXY-059 for the treatment of acute stroke: pooled analysis of the SAINT I and II Trials. Stroke. 2008;39(6):1751-8. Epub 2008 Mar 27.

Dronne MA, Grenier E, Chapuisat G, Hommel M, Boissel JP. A modelling approach to explore some hypotheses of the failure of neuroprotective trials in ischemic stroke patients. Prog Biophys Mol Biol, 2008;97(1):60-78. Epub 2007 Oct 30.

Endres M, Engelhardt B, Koistinaho J, Lindvall O, Meairs S, Mohr JP, Planas A, Rothwell N, Schwaninger M, Schwab ME, Vivien D, Wieloch T, Dirnagl U. Improving outcome after stroke: overcoming the translational roadblock. Cerebrovasc Dis, 2008;25(3):268-78. Epub 2008;22. Review.

Ford, GA. Clinical pharmacological issues in the development of acute stroke therapies. Br J Pharmacol, 2008;153 Suppl 1:S112-9. Review.

Gelmers HJ, Gorter K, de Weerdt CJ, Wiezer HJ. A controlled trial of nimodipine in acute ischemic stroke. N. Engl. J. Med, 1988;318, 203e207.

Green RA, Cross AJ. Chapter 3 Techniques for Examining Neuroprotective Drugs in Vivo International Review of Neurobiology, 199;640:47-68.

Green RA, Shuaib A. Therapeutic strategies for the treatment of stroke. Drug Discovery Today, 2006;11:681-693.

O'Collins VE, Macleod MR, Donnan GA. Horky LL, van der Worp BH, Howells DW. Experimental treatments in acute stroke. Ann. Neurol, 2006. 1,026;59, 467e477.

Ovbiagele B, Kidwell CS, Starkman S, Saver JL, Neuroprotective agents for the treatment of acute ischemic stroke. Curr. Neurol. Neurosci, 2003;Rep. 3, 9e20.

Palesch YY, Sacco RL., DeRosa JT,, Haley Jr. EC, Levin B, Ordronneau, P. Phillips SJ, Rundek T. Snipes RG, Thompson J. Glycine antagonist in neuroprotection for patientswith acute stroke:GAINAmericas: a randomized controlled trial. J. Am. Med. Assoc, 2001;285, 1719e1728.

Savitz SI, Schäbitz WR. A Critique of SAINT II: wishful thinking, dashed hopes, and the future of neuroprotection for acute stroke. Stroke, 2008;39(4):1389-91. Epub 2008;28. Review.

Scandinavian Stroke Study Group, Multicenter trial of hemodilution in acute ischemic stroke. I. Results in the total patient population. Scandinavian Stroke Study Group. Stroke, 1987;18, 691e699.

Scandinavian Stroke Study Group, Multicenter trial of hemodilution in acute ischemic stroke. Results of subgroup analyses. Scandinavian Stroke Study Group. Stroke, 1988;19, 464e471.

Serebruany VL. Hypokalemia, cardiac failure, and reporting NXY-059 safety for acute stroke. J. Cardiovasc. Pharmacol. Ther, 2006;11, 229e231.

The RANTTAS Investigators, A randomized trial of tirilazad mesylate in patients with acute stroke (RANTTAS). The RANTTAS Investigators. Stroke, 1996; 27, 1453e1458.

TRUST Study Group, Randomised, double-blind, placebo-controlled trial of nimodipine in acute stroke. Trust Study Group. Lancet, 1990;336, 1205e1209.

Yamaguchi T, Santo K, Takakura K, Saito I, Shinohara Y, Asano T, Yasuhara H. Ebselen in acute ischemic stroke: a placebo controlled, double-blind clinical trial. Ebselen Study Group. Stroke, 2000;29, 12e17.

Magnesium Sulfate in Regional and Global Cerebral ischemia:

Intravenous Magnesium Efficacy in Stroke (IMAGES) Study Investigators. Magnesium for acute stroke (Intravenous Magnesium Efficacy in Stroke Trial): randomized controlled trial. Lancet, 2004;**363**: 439–45.

Jing-Ying Lin, Shu-Ying Chung, Ming-Cheng Lin, Fu-Chou Cheng Duley, L. Effects of magnesium sulfate on energy metabolites and glutamate in the cortex during focal cerebral ischemia and reperfusion in the gerbil monitored by a dual-probe microdialysis technique Life Sciences, 2002:7 :803-811.

Meloni, B.P., Zhu, H., Knuckey, N.W. Is magnesium neuroprotective following global and focal cerebral ischaemia? A review of published studies. Magnes. Res, 2006;19:123–137.

Muir KW,., Lees KR, Ford I, Davis S. Magnesium for acute stroke (intravenous magnesium efficacy in stroke trial): randomised controlled trial. Lancet, 2004;363:439–445.

Saver JL, Kidwell C, Eckstein M, et al. Prehospital neuroprotective therapy for acute stroke: results of the field administration of stroke therapy magnesium (FAST-MAG) pilot trial. Stroke, 2004;**35**: 106–08.

Other Hypertonic Salts in Shock and Head Injury:

Rocha e Silva M, Braga GA, Prist RT, Velasco IT ,Grança ES. Isochloremic hypertonic solutions for severe hemorrhage. J Trauma, 1993;35(2):200-5.

Rocha e Silva M, Velasco IT. Hypertonic saline resuscitation: the neural component. Prog Clin Biol Res, 1989;299:303-10.

Rocha-e-Silva M, Poli de Figueiredo LF. Small volume hypertonic resuscitation of circulatory shock. Clinics, 2005;60(2):159-72. Epub 2005 Apr 26

Sheikh AA, Matsuoka T, Wisner DH. Cerebral effects of resuscitation with hypertonic saline and a new low-sodium hypertonic fluid in hemorrhagic shock and head injury. Crit Care Med, 1996;24(7):1226-32.

Velasco IT, Rocha-e-Silva M. Hyperosmotic sodium salts reverse severe hemorrhagic shock: other solutes do not. Am J Physiol, 1987;253(4 Pt 2):H751-62.

Velasco IT, Rocha-e-Silva M. Hypertonic saline resuscitation is prevented by intracerebroventricular saralasin but not by captopril. Braz J Med Biol Res, 1989;22(2):237-9.

17β-Estradiol as Neuroprotectant in Global and Regional Cerebral Ischemia:

Auriat A, Plahta WC, McGie SC, Yan R, Colbourne F. 17beta-Estradiol pretreatment reduces bleeding and brain injury after intracerebral hemorrhagic stroke in male rats. J Cereb Blood Flow Metab, 2005;25(2):247-56.

Bagetta G, Chiappetta O, Amantea D, Iannone M, Rotiroti D, Costa A, Nappi G, Corasaniti MT. Estradiol reduces cytochrome c translocation and minimizes hippocampal damage caused by transient global ischemia in rat. Neurosci Lett, 2004;16;368(1):87-91.

Beta-estradiol protects hippocampal CA1 neurons against transient forebrain ischemia in gerbil. Neurosci-Res, 1997; 29(4): 345-54.

Carswell HV, Macrae IM, Gallagher L, Harrop E, Horsburgh KJ. Neuroprotection by a selective estrogen receptor beta agonist in a mouse model of global ischemia. Am J Physiol Heart Circ Physiol, 2004;287(4):H1501-4. Epub 2004 May 20.

Dai X, Chen L, Sokabe M. Neurosteroid estradiol rescues ischemia-induced deficit in the long-term potentiation of rat hippocampal CA1 neurons. Neuropharmacology, 2007;52(4):1124-38. Epub 2007 Jan 25.

DeGiorgio LA, Attardi B, Shimizu Y, Masanori O, Volpe BT. 17β-Estradiol treatment retards excitotoxic delayed degeneration in substantia nigra reticulata neurons. Brain Research, 2002;36:15-20.

Dhandapani KM, Brann DW. Estrogen-astrocyte interactions: implications for neuroprotection. BMC Neurosci, 2002;3:6.

Dhandapani KM, Brann DW. R ole of astrocytes in estrogen-mediated neuroprotection. Exp Gerontol, 2007;42(1-2):70-5. Epub 2006 Jul 26. Review.

Gulinello M, Lebesgue D, Jover-Mengual T, Zukin RS, Etgen AM. Acute and chronic estradiol treatments reduce memory deficits induced by transient global ischemia in female rats. Horm Behav, 2006;49(2):246-60. Epub 2005 Aug 26.

He Z, He YJ, Day AL, Simpkins JW. Proestrus levels of estradiol during transient global cerebral ischemia improves the histological outcome of the hippocampal CA1 region: perfusion-dependentand-independent mechanisms. J Neurol Sci, 2002;15;193(2):79-87.

Horsburgh K, Macrae IM, Carswell H. Estrogen is neuroprotective via an apolipoprotein E-dependent mechanism in a mouse model of global ischemia.J Cereb Blood Flow Metab, 2002;22(10):1189-95.

Hurn PD, Littleton-Kearney MT, Kirsch JR, Dharmarajan AM, Traystman RJ. Postischemic cerebral blood flow recovery in the female: effect of 17 beta-estradiol.J Cereb Blood Flow Metab, 1995;15(4):666-72.

Koh PO, Cho GJ, Choi WS. 17beta-estradiol pretreatment prevents the global ischemic injury-inducedd ecrease of Akt activation and bad phosphorylation in gerbils. J Vet Med Sci, 2006;68(10):1019-22.

Littleton-Kearney MT, Gaines JM, Callahan KP, Murphy SJ, Hurn PD. Effects of estrogen on platelet reactivity after transient forebrain ischemia in rats. Biol Res Nurs, 2005;7(2):135-45.

Lu A, Ran RQ, Clark J, Reilly M, Nee A, Sharp FR. 17-beta-estradiol induces heat shock proteins in brain arteries and potentiates ischemic heat shock protein induction in glia and neurons. J Cereb Blood Flow Metab, 2002;22(2):183-95.

Miller NR, Jover T, Cohen HW, Zukin RS, Etgen AM. Estrogen can act via estrogen receptor alpha and beta to protect Hippocampal neurons against global ischemiainduced cell death. Endocrinology, 2005;146(7):3070-9. Epub 2005 Apr 7.

Qin X, Hurn PD, Littleton-Kearney MT. Estrogen restores postischemic sensitivity to the thromboxane mimetic U46619 in rat pial artery. J Cereb Blood Flow Metab, 2005;25(8):1041-6. Sandstrom NJ, Rowan MH

. Acute pretreatment with estradiol protects against CA1 cell loss and spatial learning impairments resulting from transient global ischemia. Horm Behav, 2007;51(3):335-45. Epub 2006 Dec 15.

Santizo RA, Anderson S, Ye S, Koenig HM, Pelligrino DA. Effects of estrogen on leukocyte adhesion after transient forebrain ischemia. Stroke, 2000;31(9):2231-5.

Shughrue PJ, Merchenthaler I. Estrogen prevents the loss of CA1 hippocampal neurons in gerbils after ischemic injury. Neuroscience, 2003;116(3):851-61.

Spencer SJ, Galic MA, Tsutsui M, Pittman QJ, Mouihate A. Effects of global cerebral ischemia in the pregnant rat. Stroke, 2008;39(3):975-82. Epub 2008 Jan 31.

Wang R, Zhang QG, Han D, Xu J, Lü Q, Zhang GY. Inhibition of MLK3-MKK4/7-JNK1/2 pathway by Akt1 in exogenous estrogen-induced neuroprotection against transient global cerebral ischemia by a non-genomic mechanism in male rats. J Neurochem, 2006;99(6):1543-54. Epub 2006 Oct 25.

Watanabe Y, Littleton-Kearney MT, Traystman RJ, Hurn PD. Estrogen restores postischemic pial microvascular dilation. Am J Physiol Heart Circ Physiol, 2001;281(1):H155-60.

Weigl M, Tenze G, Steinlechner B, Skhirtladze K, Reining G, Bernardo M, Pedicelli E, Dworschak M. A systematic review of currently available pharmacological neuroprotective agents as a sole intervention before anticipated or induced cardiac arrest. Resuscitation, 2005;65(1):21-39. Review.

Zhang QG, Wang R, Khan M, Mahesh V, Brann DW. Role of Dickkopf-1, an antagonist of the Wnt/beta-catenin signaling pathway, in estrogen-induced neuroprotection and attenuation of tau phosphorylation. J Neurosci, 2008;20;28(34):8430-41.

Zhou D, Matchett GA, Jadhav V, Dach N, Zhang JH. The effect of 2- thoxyestradiol, a HIF-1 alpha inhibitor, in global cerebral ischemia in rats. Neurol Res, 2008;30(3):268-71. Epub 2007 Aug 22.

Tamoxifen and Estrogen Analogs as Neuroprotectants in Global and Regional Cerebral Ischemia:

Abdullaev IF, Rudkouskaya A, Schools GP, Kimelberg HK, Mongin AA. and CI- currents in cultured rat astrocytes. J Physiol, 2006;572(Pt 3):677-89.

Bagetta G, Chiappetta O, Amantea D, Iannone M, Rotiroti D, Costa A, Nappi , Corasaniti MT. Estradiol reduces cytochrome c translocation and minimizes hippocampal damage caused by transient global ischemia in rat. Neurosci Lett, 2004;16;368(1):87-91.

Camacho A, Montiel T, Massieu L. The anion channel blocker, 4,4'dinitrostilbene-2,2'-disulfonic acid prevents neuronal death and excitatory amino acid release during glycolysis inhibition in the hippocampus in vivo.Neuroscience, 20063;142(4):1005-17. Epub 2006 Aug 22. Culmsee C, Vedder H, Ravati A, Junker V, Otto D, Ahlemeyer B, Krieg JC, Krieglstein J. Neuroprotection by estrogens in a mouse model of focal cerebral ischemia and in cultured neurons: evidence for a receptor-independent antioxidative mechanism.J Cereb Blood Flow Metab, 1999;19(11):1263-9.

Culmsee C, Vedder H, Ravati A, Junker V, Otto D, Ahlemeyer B, Krieg JC, Krieglstein J. Neuroprotection by estrogens in a mouse model of focal cerebral ischemia and in cultured neurons: evidence for a receptor-independent antioxidative mechanism. J Cereb Blood Flow Metab, 1999;19(11):1263-9.

dismutase. Endocrinology. 2008;149(1):367-79. Epub 2007 Sep 27. Ek RO, Yildiz Y, Cecen S, Yenisey C, Kavak T. Effects of tamoxifen on myocardial ischemia-reperfusion injury model in ovariectomized rats. Mol Cell Biochem, 2008;308(1-2):227-35. Epub 2007 Nov 3.

Feng Y, Fratkins JD, LeBlanc MH. Treatment with tamoxifen reduces hypoxicischemic brain injury in neonatal rats. Eur J Pharmacol, 2004;484(1):65-74.

Glembotski CC. Endoplasmic reticulum stress gene induction and protection from infarct size in MCAo and the release of glutamate in the ischemic cortical penumbra.Exp Neurol, 2008;210(2):514-20. Epub 2007 Dec 7.

Kimelberg HK, Jin Y, Charniga C, Feustel PJ. Neuroprotective activity of tamoxifen in permanent focal ischemia. J Neurosurg, 2003;99(1):138-42.

Kimelberg HK, Nestor NB, Feustel PJ. Inhibition of release of taurine and excitatory amino acids in ischemia and neuroprotection. Neurochem Res, 2004;29(1):267-74. Review.

Kimelberg HK, Nestor NB, Feustel PJ. Inhibition of release of taurine and excitatory amino acids in ischemia and neuroprotection. Neurochem Res, 2004;29(1):267-74. Review.

Kimelberg HK. Tamoxifen as a powerful neuroprotectant in experimental stroke and implications for human stroke therapy. Recent Patents CNS Drug Discov, 2008;3(2):104-8. Review.

Kimelberg HK. Astrocytic swelling in cerebral ischemia as a possible cause of injury and target for therapy. Glia, 2005;50(4):389-97. Review.

Kocic I. Modulators of ion channels activated by hypotonic swelling in cardiomyocytes: new perspectives for pharmacological treatment of life-threatening arrhythmias. Curr Med Chem Cardiovasc Hematol Agents, 2005;3(4):333-9. Review.

Ma XL, Gao F, Chen J, Christopher TA, Lopez BL, Ohlstein EH, Yue T. Endothelial protective and antishock effects of a selective estrogen receptor modulator in rats. Am J Physiol Heart Circ Physiol, 2001;280(2):H876-84.

Marcaggi P, Hirji N, Attwell D. Release of L-aspartate by reversal of glutamate transporters. Neuropharmacology, 2005;49(6):843-9. Epub 2005 Sep 16.

Martindale JJ, Fernandez R, Thuerauf D, Whittaker R, Gude N, Sussman MA, Mehta SH, Dhandapani KM, De Sevilla LM, Webb RC, Mahesh VB, Brann DW. Mendelowitsch A, Ritz MF, Ros J, Langemann H, Gratzl O. 17beta-Estradiol reduces cortical lesion size in the glutamate excitotoxicity model by enhancing extracellular lactate: a new neuroprotective pathway. Brain Res, 2001;901(1-2):230-6.

Osuka K, Feustel PJ, Mongin AA, Tranmer BI, Kimelberg HK.Tamoxifen inhibits nitrotyrosine formation after reversible middle cerebral artery occlusion in the rat. J Neurochem, 2001 Mar;76(6):1842-50.

Pharmacological comparison of swelling-activated excitatory amino acid release Phillis JW, Song D, O'Regan MH. Tamoxifen, a chloride channel blocker, reduces glutamate and aspartate release from the ischemic cerebral cortex. Brain Res, 1998;780(2):352-5.

Shughrue PJ, Merchenthaler I. Estrogen prevents the loss of CA1 hippocampal neurons in gerbils after ischemic injury. Neuroscience, 2003;116(3):851-61.

Tamoxifen neuroprotection in cerebral ischemia involves attenuation of kinase activation and superoxide production and potentiation of mitochondrial superoxide Tamoxifen, a selective estrogen receptor modulator, reduces ischemic damage caused by middle cerebral artery occlusion in the ovariectomized female rat. Neuroendocrinology, 2003;77(1):44-50.

Zhang Y, Jin Y, Behr MJ, Feustel PJ, Morrison JP, Kimelberg HK. Behavioral and histological neuroprotection by tamoxifen after reversible focal cerebral ischemia.Exp Neurol, 2005;196(1):41-6. Epub 2005 Jul 28.

Zhang Y, Milatovic D, Aschner M, Feustel PJ, Kimelberg HK. Neuroprotection by tamoxifen in focal cerebral ischemia is not mediated by an agonist action at estrogen receptors but is associated with antioxidant activity. Exp Neurol, 2007;204(2):819-27. Epub 2007 Jan 24.

Failure of Nimodipine in Clinical Trials for Post-Ischemic Encephalopathy:

Forsman M, Aarseth HP, Nordby HK, Skulberg A, Steen PA. Effects of nimodipine on cerebral blood flow and cerebrospinal fluid pressure after cardiac arrest: correlation with neurologic outcome. Anesth Analg, 1989;68(4):436-43.

Gueugniaud PY, Gaussorgues P, Garcia-Darennes F, Bancalari G, Roux H, Robert D, Petit P. Early effects of nimodipine on intracranial and cerebral perfusion pressures in cerebral anoxia after out-of-hospital cardiac arrest. Resuscitation, 1990;20(3):203-12.

Huyghens LP, Buylaert WA, Corne L, Rosseel MT, Bogaert MG. Plasma concentrations and haemodynamic effects of nimodipine in patients resuscitated after cardiac arrest. Eur J Clin Pharmacol, 1989;36(4):327-33.

Prough DS, Furberg CD. Nimodipine and the "no-reflow phenomenon"--experimental triumph, clinical failure? Anesth Analg, 1989;68(4):431-5.

Roine RO, Kajaste S, Kaste M. Neuropsychological sequelae of cardiac arrest. JAMA, 1993 Jan 13;269(2):237-42.

Roine RO, Kaste M, Kinnunen A, Nikki P, Sarna S, Kajaste S. Nimodipine after resuscitation from out-of-hospital ventricular fibrillation. A placebo-controlled, double-blind, randomized trial. JAMA, 1990;264(24):3171-7.

Roine RO, Kaste M, Kinnunen A, Nikki P. Safety and efficacy of nimodipine in resuscitation of patients outside hospital.Br Med J (Clin Res Ed), 987;294(6563):20.